

				Effective	date: 01/02/202	4			
OPERATIVE DETAILS						Section			
Donor Hospital			NRP Centre						
Name			NRP Centre Contact Number						
DoB			Cardiothoracic Retrieval (circle)	NO / LUN	NO / LUNG / HEART / TA NRP				
NHS or CHI No	HI No		Cardiothoracic Team(s))					
			CT Lead Surgeon (PRINT)	ı (PRINT)					
ODT Donor No			CT Organ Preservation						
Blood Group			Stapler technique (lungs)	YES / N	0				
Height (cm) / Weight (kg) / Girth (cm)			NRP Perfusion Specialist	t					
Donor allergies			AB Organ Preservation	ı					
SNOD			NRP Surgeon (PRINT))					
SNOD contact no			NRP Surgeon contact no)					
			NRP Surgeon signature			_			
	By signing here, the	e NRP surgeo	n is prescribing all drugs, fluids, an	nd blood prod	lucts as initialle	d on chart			
FLUIDS/EQUIPMENT						Section			
PRIME									
	Amount/Vol	Expiry	Batch/Ref (DIN)	Rx	Checked	Given			
Sterile Circuit									
Hartmann's Solution	1000ml								
Hartmann's Solution	1000ml								
Sodium Bicarbonate 8.4% (1ml/kg)									
Heparin	50,000 units								
Fluconazole 2mg/mL	400mg								
Teicoplanin	200mg								
Gentamicin	120mg								
Metronidazole	500mg								
Methylprednisolone	1g								
Phentolamine	5mg								
ADDITIONAL FLUIDS									
Blood									
Blood									
Blood									
Blood									
						1			
						1			
						1			
		1							

ODT Donor No

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			Section 3
Femoral / EIA / CIA / Aorta	Arterial cannula size		
Femoral / EIV / CIV / IVC	Venous cannula size		
Mandatory prior to NRP start	Vent cannula size		
External CLAMP	IVC clamp on (circle)	YES / NO	
Endoclamp (Scotland only)	SVC clamp on (circle)	YES / NO	
	Femoral / EIV / CIV / IVC Mandatory prior to NRP start External CLAMP	Femoral / EIV / CIV / IVC Venous cannula size Mandatory prior to NRP start External CLAMP IVC clamp on (circle)	Femoral / EIV / CIV / IVC Venous cannula size Mandatory prior to NRP start External CLAMP IVC clamp on (circle) YES / NO

IP PARAME	TERS							Secti
lock ime NRP Duration	Blood Flow	FiO ₂	Gas Flow (I/min)	SvO ₂ %	HCT %	Reservoir Volume	Temp °C	Notes
0								
10								
20								
30								
40								
50								
60								
70								
80								
90								
100								
110								
120								



ASES	SES Section										
Clock time											
NRP duration	0	30		60	90	120					
Sample type (A/V)											
рН											
pCO ₂											
pO ₂											
HCO ₃											
BE											
Sats											
Lact											
Na⁺											
K⁺											
Gluc											
Ca ²⁺											
Hct											
Hb											

				1	-
Clock time					
NRP duration 0	30	60	90	120	
Gluc					
Urea					
Crea					
Uric Acid					
Ca					
ALB					
Tot Prot					
ALT					
AST					
ALP					
Bili					
GGT					
AMY					

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MINGS								Section 7
VLST location (circle) ITU / THEATRE SUITE				Knife	to skin			
WLST				Aortic arch	vented			
stolic BP < 50 mmHg				IRP Start (T	ime 0)			
Asystole				NRP sto	p time			
Verified Deceased				In situ colo	l Flush			
ORGAN QUALITY ASSESSMENT								Section 8
Liver weight kg Q Minimum dataset: LFT/amylase is 0 , 60 and 120 minutes. Glucose, lactate and haemoglobin data a Blood cultures may be taken at 0 and 120		ed at 0, 30		nd 120 mi		IRP Box No		
	0	30	60	90	120	Validating Lab	Notes	
ALT (U/I)								
BILI (umol/l)								
AMYLASE (U/I)								
GLUCOSE (mmol/l)								
LACTATE (mmol/l)								
Hb (g/l)								
BLOOD CULTURES (optional)								

INSTRUCTIONS FOR SNODS: Take a copy of pages 1-4 on iPad via genius scan app and email to anrp_passport_alert@nhsbt.nhs.uk to ensure receipt by all required.

Guidance Notes for the NRP Passport and NRP Management (FRM6725)

Section 2. Fluids/Equipment

Hartmann's solution is used in the Prime. Blood may also be used in the prime for paediatric donors, in TANRP or if starting haemoglobin is low. Hartmann's contains high levels of lactate (27mM) and if added after pump start, will lead to confusion regarding lactate metabolism by the liver.

Avoid further Hartmann's solution once NRP has commenced as it contains lactate.

Section 5. Gases

Arterial blood gases are essential to ensure appropriate acid-base status, tissue oxygenation and CO_2 levels and best describe oxygenator function and acid base status. Repeated arterial blood gases are **strongly recommended** for safe perfusion in NRP. Please ensure that gas samples are capped immediately to prevent loss of CO_2 .

pH and Bicarbonate. pH<7 or H+>100nmol/l at time 0 can lead to mitochondrial dysfunction. Therefore add 25ml (or 50ml) 8.4% NaHCO₃ once, as early as possible in this situation. The resolution of acidosis during NRP is an index of organ quality so adding more bicarbonate later will be confusing.

Avoid further Bicarbonate administration once NRP is in progress.

PaCO₂. Optimise arterial CO₂ by adjusting gas flow rate. If PaCO₂ is <4.5kPa, decrease gas flow rate; if paCO₂>6.0 kPa, increase gas flow rate.

Optimise PaO₂ by changing FiO2 alone.

Venous blood gases give an accurate measurement of SvO_2 . Mixed venous oxygen saturation (SvO_2) is an essential measure of tissue oxygen uptake, and hence adequacy of oxygen delivery to tissues. However, other analytes in venous gases (pH, pCO₂ etc) cannot be used to gauge acid-base status or gas exchange accurately, as normal venous ranges are not defined. This is why arterial blood gases are recommended for safe perfusion.

SvO₂ between 60% and 80% indicates satisfactory oxygen delivery (good flow, good arterial O₂, adequate Hb) to tissues.

Optimise SvO_2 by adjusting pump flow rate. If $SvO_2 < 60\%$, increase pump flow; if $SvO_2 > 80\%$ decrease pump flow (NB; Hb needs to be >60g/l for SvO_2 to be reliable).

When interpreting arterial gases, at least one set of venous gases should be performed for to confirm the SvO₂ monitor on the perfusion device is reading correctly. They tend not to read well if Hb is low, which may trigger transfusion to promote adequate oxygen delivery.

Section 8. Organ Quality Assessment Bloods.

The 'Validating Lab' is the NHS Laboratory which quality assures the point-of-care-testing ('POCT') device which is used in the donor theatre for rapid blood results. This will either be the donor hospital lab, for bloods tested with donor hospital equipment, or the NORS base hospital lab which has provided quality assurance for the POCT devices used by the NORS team, or both.

LFTs and gases are processed with POCT devices ('Piccolo', 'i-Stat') or via the Donor Hospital Lab. Haemoglobin and lactate are measured on the gas sample. Whichever device/laboratory is used, please ensure that the same device/laboratory/sample type is used on each occasion.

Red cells in a gas sample may settle to the bottom of a sample tube, and cause inaccurate Hb readings. Please ensure gas samples are well mixed prior to analysis.