

A thick blue wavy line that curves across the top of the slide, separating the header from the main title.

Crossmatching strategies prior to deceased donor kidney transplantation

Chris Callaghan

Abdominal Organ Utilisation Lead, NHSBT

Tracey Rees

**Head of the Welsh Transplantation and
Immunogenetics Laboratory, KAG BSHI Rep**

Clinical case

- June 2018 – ‘ideal’ DBD kidney donor aged in their 40’s
- 1200 – cross-clamp; organ then transported 100 miles away
- 2300 – implanting unit reported a positive crossmatch on LN / spleen
 - Policy of using PBLs for XM, but only from ‘local’ donors (due to cost concerns)
- Re-offered to child at another centre; declined due to prolonged CIT
- Declined by 3 other centres in KFTS due to prolonged CIT
- Eventually accepted and implanted by fourth centre

Background

- No current UK guidance on optimal XM strategies prior to deceased donor kidney transplantation
 - PBL XM in live donor programmes; increasing use of virtual XM (VXM)
- Negative PBL XM / VXM; reduces CIT by 2.5-3.5 hours in the UK
 - Taylor CJ, *Transplantation* 2010
 - Shrestha S, *Transplantation* 2016
- Positive PBL XM / VXM; optimal offering pathway and likely to reduce the risk of discard due to prolonged CIT

Background

- Costs:
 - Transporting PBLs 100 miles: £210 (Amvale)
 - 24-hour inpatient stay one day: £215 (Guy's Hospital)
 - One session of inpatient dialysis: £150 (Guy's Hospital)
- National survey of H+I laboratories
- KAG discussion
 - Support for national guidance via BSHI/BTS
 - Aim to avoid waiting for LN/spleen XM in deceased donor kidney transplantation
- **Update renal transplant community with planned developments**

Compatibility tests pre-transplant

Strategy / source	Available prior to organ transport?	Timing	Technical	Logistical issues
Wet XM Splenic derived lymphocytes	No	4-6 hours after delivery to laboratory	Usually plenty of starting material. Good viability	Can be transported at same time as organs
Wet XM Peripheral blood lymphocytes	Yes – dependent on timings and logistics	4-6 hours after delivery to laboratory	Perceived quality/sensitivity issues.	Needs separate transport to more than one centre Transport cost
Virtual crossmatching	Yes	1 hour if using historic test results; 3 hours if fresh testing required	Detailed donor typing needed. Knowledge-based risk assessment for sensitised patients	May require collection / testing of fresh recipient sample

BSHI survey

- Current practices for deceased donor crossmatching
 - Local vs imported
 - Cell source (PBL / spleen / lymph nodes)
- Use of VXM (sensitised and unsensitised)
- Out-of-hours antibody testing
- Turn-around times

BSHI survey – results

- 17/21 laboratories responded
- Results available 4-6 hours
- 5/17 (29%) routinely use PBL XM
- 11/17 use PBL XM for ‘local’ donors and 7 of these sometimes use PBL for imported donors
- Only one laboratory NEVER uses PBL XM

BSHI survey – results

- Reasons for not performing PBL XM:
 - Concerns about quality and number of lymphocytes
 - Transport costs
 - Not beneficial as 85% of transplants proceed with VXM
 - Didn't know it was an option for imported donors (one laboratory)
- VXM
 - 13/17 (76%) routinely use VXM for all unsensitised patients, with remaining 4 using VXM sometimes
 - 14/17 use for sensitised patients
 - 11/17 perform HLA Ab testing out-of-hours to support VXM, if required

Conclusions

- Variation in both clinical / H+I practices
- National guidance supported by KAG / BSHI
- Next steps:
 - Workshop 22 March, Birmingham
 - Reps from H+I labs / clinical teams
 - Shared practices, evidence-based
 - Agree and publish national BSHI / BTS guidelines for deceased donor kidney crossmatching
 - Draft by KAG June 2019
 - **Aim to have crossmatch results before organs are removed**

Acknowledgements

KAG

Chris Watson
Sue Fuggle
Natalia Diaz Burlinson

NHSBT

Jenny Mehew
Winter Hughes
Anthony Clarkson
Andrea Harmer

BSHI

David Briggs

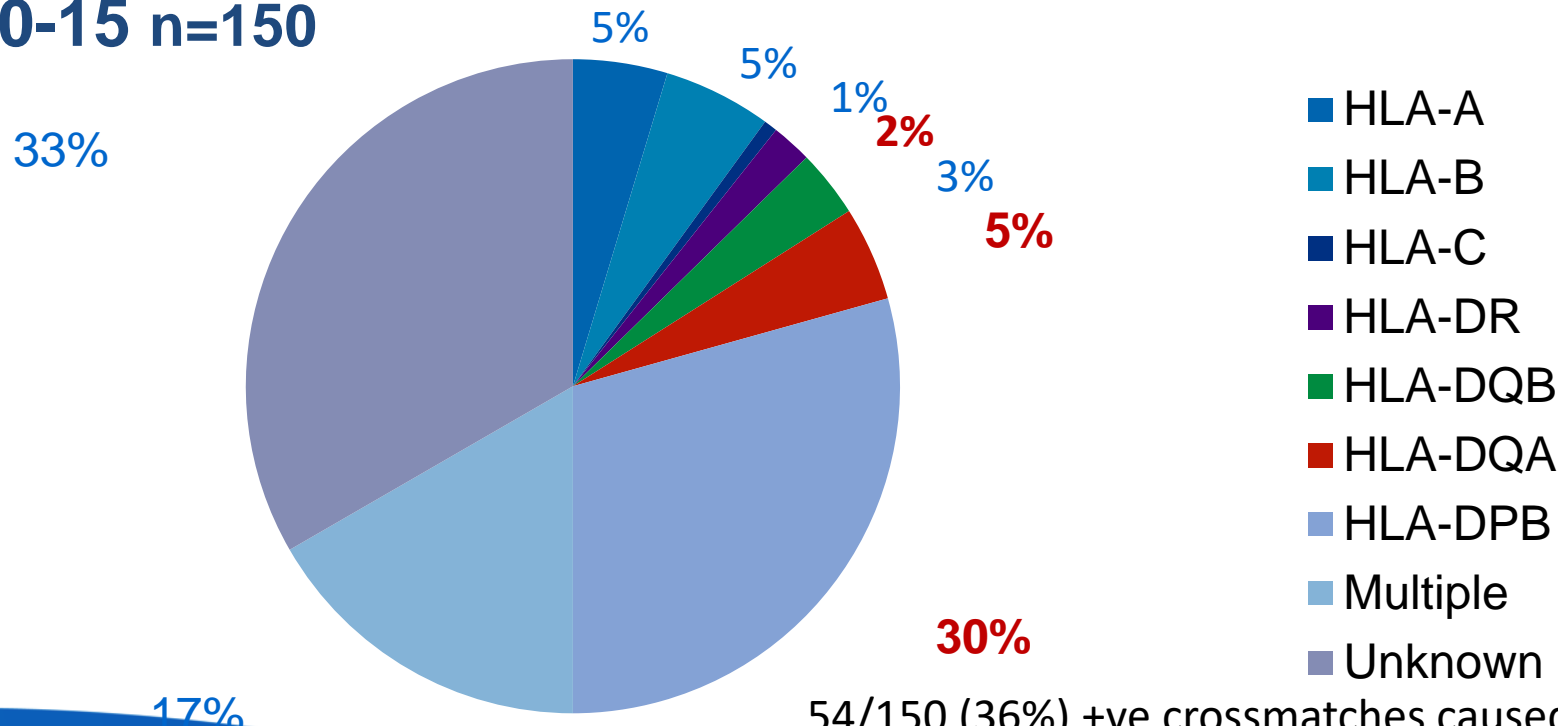
Kidneys Allocated: 2010-2015

Positive crossmatches

Year	Kidneys Allocated	Positive crossmatch n=	%
2010	976	36	3.7
2011	938	26	2.7
2012	956	23	2.4
2013	1138	25	2.2
2014	1180	24	2.0
2015	1112	16	1.4
Total	6300	150	2.4

Reasons for a Positive Crossmatch:

2010-15 n=150



54/150 (36%) +ve crossmatches caused by specificities, DP, DQA and some DR alleles, outside the required minimum resolution